

Order information

REF	CONTENT	Analyzer(s) on which cobas c pack(s) can be used
03507343 190	Tina-quant IgA Gen.2 150 tests	System-ID 07 6786 7 Roche/Hitachi cobas c 311, cobas c 501/502
11355279 216	Calibrator f.a.s. Proteins (5 x 1 mL)	Code 656
11355279 160	Calibrator f.a.s. Proteins (5 x 1 mL, for USA)	Code 656
10557897 122	Precinorm Protein (3 x 1 mL)	Code 302
10557897 160	Precinorm Protein (3 x 1 mL, for USA)	Code 302
11333127 122	Precipath Protein (3 x 1 mL)	Code 303
11333127 160	Precipath Protein (3 x 1 mL, for USA)	Code 303
10171743 122	Precinorm U (20 x 5 mL)	Code 300
10171735 122	Precinorm U (4 x 5 mL)	Code 300
03121291 122	Precipath PUC (4 x 3 mL)	Code 241
05117003 190	PreciControl ClinChem Multi 1 (20 x 5 mL)	Code 391
05947626 190	PreciControl ClinChem Multi 1 (4 x 5 mL)	Code 391
05947626 160	PreciControl ClinChem Multi 1 (4 x 5 mL, for USA)	Code 391
05117216 190	PreciControl ClinChem Multi 2 (20 x 5 mL)	Code 392
05947774 190	PreciControl ClinChem Multi 2 (4 x 5 mL)	Code 392
05947774 160	PreciControl ClinChem Multi 2 (4 x 5 mL, for USA)	Code 392
04489357 190	Diluent NaCl 9 % (50 mL)	System-ID 07 6869 3

English

System information

For **cobas c** 311/501 analyzers:

IGA-2: ACN 458 (Standard application)

IGAP2: ACN 295 (Sensitive application)

For **cobas c** 502 analyzer:

IGA-2: ACN 8458 (Standard application)

IGAP2: ACN 8295 (Sensitive application)

Intended use

In vitro test for the quantitative determination of IgA in human serum and plasma on Roche/Hitachi **cobas c** systems.

Summary^{1,2,3,4,5,6,7,8,9,10,11,12}

IgA accounts for 13 % of the plasma immunoglobulins and serves to protect the skin and mucosa against microorganisms. It is capable of binding toxins, and in combination with lysozyme develops anti-bacterial and antiviral activity. IgA is the predominant immunoglobulin in bodily secretions such as colostrum, saliva and sweat. Secretory IgA provides defense against local infections and is important in binding food antigens in the gut. In serum, IgA exists in monomeric, dimeric and trimeric forms, whereas in bodily secretions it exists exclusively in dimeric form with an additional chain (secretory component).

Increased polyclonal IgA levels may occur in chronic liver diseases, chronic infections, autoimmune disorders (rheumatoid arthritis, systemic lupus erythematosus), sarcoidosis and Wiscott-Aldrich syndrome. Monoclonal IgA increases in IgA myeloma.

Decreased synthesis of IgA is observed in acquired and congenital immunodeficiency diseases such as Bruton type agammaglobulinemia. Reduced levels of IgA can be caused by protein-losing gastroenteropathies and loss through skin from burns.

Due to the slow onset of IgA synthesis, the IgA concentration in serum of infants is lower than in adults.

Use of specific antibodies for quantitation of serum proteins has become a valuable diagnostic tool. Light-scattering properties of antigen/antibody aggregates were first observed by Pope and Healey in 1938, and later confirmed by Gitlin and Edelhoch. Ritchie employed turbidimetric measurements to quantitate specific proteins. Quantitation of immunoglobulins can also be done using nephelometric techniques. Polymeric enhancement with polyethylene glycol (PEG) to improve sensitivity and increase the rate of antigen/antibody complex formation has been described by Lizana and Hellsing.

The Roche IgA assay is based on the principle of immunological agglutination.

In addition to the standard application (test IGA-2), there is a sensitive application (test IGAP2) designed for the quantitative determination of low IgA concentrations, e.g. in pediatric samples.

It is known that the so-called paraproteins secreted in monoclonal gammopathies (monoclonal immunoglobulinemia) may differ from the respective immunoglobulins of polyclonal origin by amino acid composition and size. This may impair the binding to antibody and hence impair accurate quantitation.

Test principle

Immunoturbidimetric assay

Anti-IgA antibodies react with antigen in the sample to form an antigen/antibody complex. Following agglutination, this is measured turbidimetrically. Addition of PEG allows the reaction to progress rapidly to the end point, increases sensitivity, and reduces the risk of samples containing excess antigen producing false negative results.

Reagents - working solutions

R1 TRIS buffer: 20 mmol/L, pH 8.0; NaCl: 200 mmol/L; polyethylene glycol: 3.6 %; preservative; stabilizers

R2 Anti-human IgA antibody (goat): dependent on titer; TRIS buffer: 20 mmol/L, pH 8.0; NaCl: 150 mmol/L; preservative

R1 is in position B and R2 is in position C.

Precautions and warnings

For in vitro diagnostic use.

Exercise the normal precautions required for handling all laboratory reagents.

Disposal of all waste material should be in accordance with local guidelines. Safety data sheet available for professional user on request.

Reagent handling

Ready for use

Storage and stability

IGA-2

Shelf life at 2-8 °C:

See expiration date on **cobas c** pack label.

On-board in use and refrigerated on the analyzer:

12 weeks

Diluent NaCl 9 %

Shelf life at 2-8 °C:

See expiration date on **cobas c** pack label.

IGA-2

Tina-quant IgA Gen.2



On-board in use and refrigerated on the analyzer: 12 weeks

Specimen collection and preparation

For specimen collection and preparation only use suitable tubes or collection containers.

Only the specimens listed below were tested and found acceptable.

Standard application (IGA-2)

Serum

Plasma: Li-heparin and K₂-EDTA plasma

Sensitive application (IGAP2)

Serum

Plasma: Li-heparin and K₂-EDTA plasma

The sample types listed were tested with a selection of sample collection tubes that were commercially available at the time of testing, i.e. not all available tubes of all manufacturers were tested. Sample collection systems from various manufacturers may contain differing materials which could affect the test results in some cases. When processing samples in primary tubes (sample collection systems), follow the instructions of the tube manufacturer.

Partially filled K₂-EDTA plasma tubes can cause incorrect results.

Centrifuge samples containing precipitates before performing the assay.

Stability:¹³ 8 months at 15-25 °C
8 months at 2-8 °C
8 months at (-15)-(-25) °C

Materials provided

See "Reagents – working solutions" section for reagents.

Materials required (but not provided)

See "Order information" section

General laboratory equipment

Assay

For optimum performance of the assay follow the directions given in this document for the analyzer concerned. Refer to the appropriate operator's manual for analyzer-specific assay instructions.

The performance of applications not validated by Roche is not warranted and must be defined by the user.

Application for serum and plasma

Standard application (IGA-2)

cobas c 311 test definition

Assay type	2-Point End		
Reaction time / Assay points	10 / 6-31		
Wavelength (sub/main)	700/340 nm		
Reaction direction	Increase		
Units	g/L (μmol/L, mg/dL)		
Reagent pipetting	Diluent (H ₂ O)		
R1	120 μL	–	
R2	38 μL	–	
Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	5 μL	9 μL	180 μL
Decreased	2.7 μL	2 μL	180 μL
Increased	2.4 μL	–	–

cobas c 501/502 test definition

Assay type	2-Point End
Reaction time / Assay points	10 / 10-46
Wavelength (sub/main)	700/340 nm

Reaction direction	Increase		
Units	g/L (μmol/L, mg/dL)		
Reagent pipetting	Diluent (H ₂ O)		
R1	120 μL	–	
R2	38 μL	–	
Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	5 μL	9 μL	180 μL
Decreased	2.7 μL	2 μL	180 μL
Increased	2.4 μL	–	–

Sensitive application (IGAP2)

cobas c 311 test definition

Assay type	2-Point End		
Reaction time / Assay points	10 / 6-22		
Wavelength (sub/main)	700/340 nm		
Reaction direction	Increase		
Units	g/L (μmol/L, mg/dL)		
Reagent pipetting	Diluent (H ₂ O)		
R1	120 μL	–	
R2	38 μL	–	
Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	10 μL	9 μL	75 μL
Decreased	7 μL	5 μL	93 μL
Increased	2.7 μL	–	–

cobas c 501/502 test definition

Assay type	2-Point End		
Reaction time / Assay points	10 / 10-46		
Wavelength (sub/main)	700/340 nm		
Reaction direction	Increase		
Units	g/L (μmol/L, mg/dL)		
Reagent pipetting	Diluent (H ₂ O)		
R1	120 μL	–	
R2	38 μL	–	
Sample volumes	Sample	Sample dilution	
		Sample	Diluent (NaCl)
Normal	10 μL	9 μL	75 μL
Decreased	7 μL	5 μL	93 μL
Increased	2.7 μL	–	–

Calibration

Standard application (IGA-2)

Calibrators	S1: H ₂ O S2-S6: C.f.a.s. Proteins	
	Multiply the lot-specific C.f.a.s. Proteins calibrator value by the factors below to determine the standard concentrations for the six-point calibration curve:	
	S2: 0.100	S5: 1.00
	S3: 0.250	S6: 2.00
	S4: 0.501	

Calibration mode	RCM
Calibration frequency	Full calibration - after reagent lot change - as required following quality control procedures

Sensitive application (IGAP2)

Calibrators	S1: H ₂ O S2-S6: C.f.a.s. Proteins
	Multiply the lot-specific C.f.a.s. Proteins calibrator value by the factors below to determine the standard concentrations for the six-point calibration curve:
	S2: 0.0205 S5: 0.252
	S3: 0.0609 S6: 1.00
	S4: 0.126

Calibration mode	RCM
Calibration frequency	Full calibration - after reagent lot change - as required following quality control procedures

Traceability: This method has been standardized against the reference preparation of the IRMM (Institute for Reference Materials and Measurements) BCR470/CRM470 (RPPHS - Reference Preparation for Proteins in Human Serum).¹⁴

Quality control

For quality control, use control materials as listed in the "Order Information" section. In addition, other suitable control material can be used.

Standard application (IGA-2):	Precinorm Protein, Precipath Protein, Precinorm U, PreciControl ClinChem Multi 1, PreciControl ClinChem Multi 2
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Sensitive application (IGAP2):	Precinorm Protein, Precipath PUC, PreciControl ClinChem Multi 1
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The control intervals and limits should be adapted to each laboratory's individual requirements. Values obtained should fall within the defined limits. Each laboratory should establish corrective measures to be taken if values fall outside the defined limits.

Follow the applicable government regulations and local guidelines for quality control.

Calculation

Roche/Hitachi **cobas c** systems automatically calculate the analyte concentration of each sample.

Conversion factors:	mg/dL x 0.01 = g/L	g/L x 6.25 = μmol/L
	g/L x 100 = mg/dL	μmol/L x 0.16 = g/L

Limitations - interference**Standard application (IGA-2):**

Criterion: Recovery within ± 10 % of initial value at an IgA concentration of 0.70 g/L (4.38 μmol/L, 70 mg/dL).

Icterus:¹⁵ No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026 μmol/L or 60 mg/dL).

Hemolysis:¹⁵ No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 621 μmol/L or 1000 mg/dL).

Lipemia (Intralipid):¹⁵ No significant interference up to an L index of 2000. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Rheumatoid factors < 1200 IU/mL do not interfere.

High dose hook-effect: No false result up to an IgA concentration of 100 g/L (625 μmol/L, 10000 mg/dL) occurs due to an antigen excess within polyclonal specimens.

There is no cross-reaction between IgA and IgG or IgM under the assay conditions.

Drugs: No interference was found at therapeutic concentrations using common drug panels.^{16,17}

Sensitive application (IGAP2):

Criterion: Recovery within ± 10 % of initial value at an IgA concentration of 0.40 g/L (2.5 μmol/L, 40 mg/dL).

Icterus:¹⁵ No significant interference up to an I index of 60 for conjugated and unconjugated bilirubin (approximate conjugated and unconjugated bilirubin concentration: 1026 μmol/L or 60 mg/dL).

Hemolysis:¹⁵ No significant interference up to an H index of 1000 (approximate hemoglobin concentration: 621 μmol/L or 1000 mg/dL).

Lipemia (Intralipid):¹⁵ No significant interference up to an L index of 2000. There is poor correlation between the L index (corresponds to turbidity) and triglycerides concentration.

Rheumatoid factors < 500 IU/mL do not interfere.

High dose hook-effect: No false result up to an IgA concentration of 20 g/L (125 μmol/L, 2000 mg/dL) occurs due to an antigen excess within polyclonal specimens.

There is no cross-reaction between IgA and IgG or IgM under the assay conditions.

Drugs: No interference was found at therapeutic concentrations using common drug panels.^{16,17}

As with other turbidimetric or nephelometric procedures, this test may not provide accurate results in patients with monoclonal gammopathy, due to individual sample characteristics which can be assessed by electrophoresis.¹⁸

For diagnostic purposes, the results should always be assessed in conjunction with the patient's medical history, clinical examination and other findings.

ACTION REQUIRED

Special Wash Programming: The use of special wash steps is mandatory when certain test combinations are run together on Roche/Hitachi **cobas c** systems. The latest version of the carry-over evasion list can be found with the NaOHD/SMS/Multiclean/SCCS or the NaOHD/SMS/SmpCln1+2/SCCS Method Sheets. For further instructions refer to the operator's manual. **cobas c** 502 analyzer: All special wash programming necessary for avoiding carry-over is available via the **cobas** link, manual input is not required.

Where required, special wash/carry-over evasion programming must be implemented prior to reporting results with this test.

Limits and ranges**Measuring range****Standard application (IGA-2):**

0.50-8.00 g/L (3.13-50 μmol/L, 50-800 mg/dL)

Determine samples having higher activities via the rerun function. Dilution of samples via the rerun function is a 1:8 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 8.

Determine samples having lower concentrations via the rerun function. For samples with lower concentrations, the rerun function increases the sample volume by a factor of 10. The results are automatically divided by this factor.

Sensitive application (IGAP2):

0.1-4.0 g/L (0.63-25 μmol/L, 10-400 mg/dL)

Determine samples having higher activities via the rerun function. Dilution of samples via the rerun function is a 1:3 dilution. Results from samples diluted using the rerun function are automatically multiplied by a factor of 3.

Determine samples having lower concentrations via the rerun function. For samples with lower concentrations, the rerun function increases the sample volume by a factor of 2.5. The results are automatically divided by this factor.

Lower limits of measurement**Standard application (IGA-2):****Limit of Blank (LoB) and Limit of Detection (LoD)**

Limit of Blank	= 0.05 g/L
Limit of Detection	= 0.05 g/L

The Limit of Blank and Limit of Detection were determined in accordance with the CLSI (Clinical and Laboratory Standards Institute) EP17-A requirements.

The Limit of Blank is the 95th percentile value from $n \geq 60$ measurements of analyte-free samples over several independent series. The Limit of Blank corresponds to the concentration below which analyte-free samples are found with a probability of 95 %.

The Limit of Detection is determined based on the Limit of Blank and the standard deviation of low concentration samples.

The Limit of Detection corresponds to the lowest analyte concentration which can be detected (value above the Limit of Blank with a probability of 95 %).

Sensitive application (IGAP2):

Lower detection limit of the test

0.04 g/L (0.25 $\mu\text{mol/L}$, 4 mg/dL)

The lower detection limit represents the lowest measurable analyte level that can be distinguished from zero. It is calculated as the value lying 3 standard deviations above that of the lowest standard (standard 1 + 3 SD, repeatability, $n = 21$).

Expected values

Reference values according to CRM 470 Protein Standardization:^{19,20}

Adults	0.7-4 g/L	4.38-25.0 $\mu\text{mol/L}$	70-400 mg/dL
Children and juveniles			
0-1 year	0.00-0.83 g/L	0.00-5.19 $\mu\text{mol/L}$	0.00-83 mg/dL
1-3 years	0.20-1.00 g/L	1.25-6.25 $\mu\text{mol/L}$	20-100 mg/dL
4-6 years	0.27-1.95 g/L	1.69-12.19 $\mu\text{mol/L}$	27-195 mg/dL
7-9 years	0.34-3.05 g/L	2.13-19.06 $\mu\text{mol/L}$	34-305 mg/dL
10-11 years	0.53-2.04 g/L	3.31-12.75 $\mu\text{mol/L}$	53-204 mg/dL
12-13 years	0.58-3.58 g/L	3.63-22.38 $\mu\text{mol/L}$	58-358 mg/dL
14-15 years	0.47-2.49 g/L	2.94-15.56 $\mu\text{mol/L}$	47-249 mg/dL
16-19 years	0.61-3.48 g/L	3.81-21.75 $\mu\text{mol/L}$	61-348 mg/dL

Roche has not evaluated reference ranges in a pediatric population.

Each laboratory should investigate the transferability of the expected values to its own patient population and if necessary determine its own reference ranges.

Specific performance data

Representative performance data on the analyzers are given below. Results obtained in individual laboratories may differ.

Precision

Precision was determined using human samples and controls in an internal protocol with repeatability ($n = 21$) and intermediate precision (3 aliquots per run, 1 run per day, 21 days). The following results were obtained:

Standard application (IGA-2):

<i>Repeatability</i>	<i>Mean</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>SD</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>CV</i> %
Precinorm Protein	1.95 (12.2, 195)	0.02 (0.1, 2)	1.1
Precipath Protein	3.23 (20.2, 323)	0.02 (0.1, 2)	0.7
Human serum 1	1.55 (9.69, 155)	0.02 (0.13, 2)	1.0
Human serum 2	2.23 (13.9, 223)	0.02 (0.1, 2)	0.9
<i>Intermediate precision</i>	<i>Mean</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>SD</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>CV</i> %
Precinorm Protein	1.95 (12.2, 195)	0.03 (0.2, 3)	1.8
Precipath Protein	3.25 (20.3, 325)	0.04 (0.3, 4)	1.4
Human serum 3	1.93 (12.1, 193)	0.04 (0.3, 4)	1.8
Human serum 4	3.31 (20.7, 331)	0.04 (0.3, 4)	1.1

Sensitive application (IGAP2):

<i>Repeatability</i>	<i>Mean</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>SD</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>CV</i> %
Precipath PUC	0.27 (1.69, 27.0)	0.01 (0.06, 1.0)	1.8
Precinorm Protein	2.24 (14.0, 224)	0.02 (0.1, 2)	0.9
Human serum 1	0.37 (2.31, 37.0)	0.01 (0.06, 1.0)	1.3
Human serum 2	2.40 (15.0, 240)	0.02 (0.1, 2)	0.8
<i>Intermediate precision</i>	<i>Mean</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>SD</i> g/L ($\mu\text{mol/L}$, mg/dL)	<i>CV</i> %
Precipath PUC	0.27 (1.69, 27.0)	0.01 (0.06, 1.0)	3.2
Precinorm Protein	2.25 (14.1, 225)	0.04 (0.3, 4)	1.8
Human serum 3	0.36 (2.25, 36.0)	0.01 (0.06, 1.0)	2.4
Human serum 4	1.26 (7.89, 126)	0.02 (0.13, 2)	1.5

Method comparison

IgA values for human serum and plasma samples obtained on a Roche/Hitachi **cobas c 501** analyzer (y) were compared with those determined using the same reagent on a Roche/Hitachi 917 analyzer (x).

Standard application (IGA-2):

Sample size (n) = 79

Passing/Bablok ²¹	Linear regression
$y = 1.035x - 0.019$ g/L	$y = 1.027x - 0.003$ g/L
$r = 0.987$	$r = 0.999$

The sample concentrations were between 0.500 and 7.74 g/L (3.13 and 48.4 $\mu\text{mol/L}$, 50.0 and 774 mg/dL).

Sensitive application (IGAP2):

Sample size (n) = 194

Passing/Bablok ²¹	Linear regression
$y = 0.981x + 0.002$ g/L	$y = 0.956x + 0.035$ g/L
$r = 0.957$	$r = 0.998$

The sample concentrations were between 0.166 and 4.00 g/L (1.06 and 25.0 $\mu\text{mol/L}$, 16.6 and 400 mg/dL).

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A point (period/stop) is always used in this Method Sheet as the decimal separator to mark the border between the integral and the fractional parts of a decimal numeral. Separators for thousands are not used.

Symbols

Roche Diagnostics uses the following symbols and signs in addition to those listed in the ISO 15223-1 standard.

	Contents of kit
	Volume after reconstitution or mixing

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